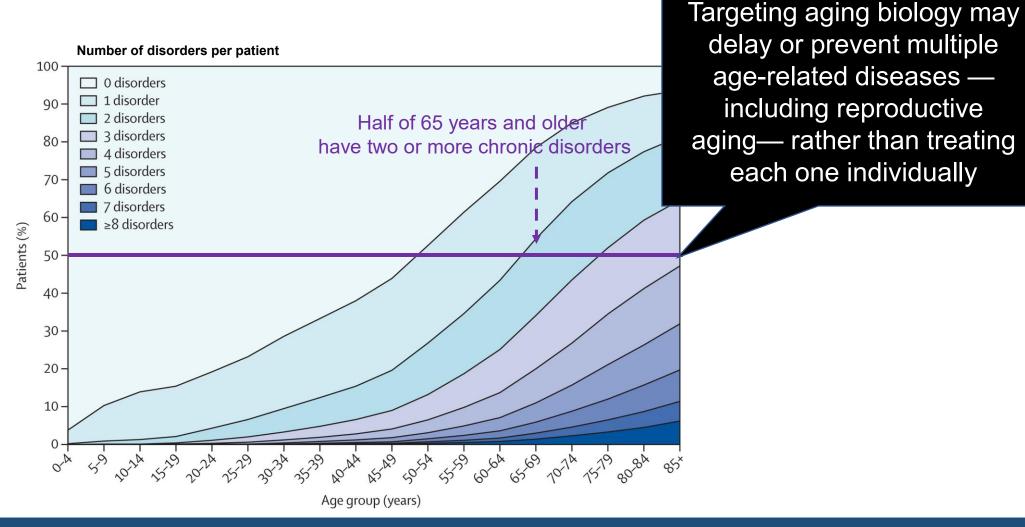


Unlocking Female Reproductive Longevity : The Role of Cellular Senescence in Ovarian Aging

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Aging as a root cause of age-related chronic diseases



Aging is the primary risk factor for multimorbidity and chronic diseases

Why ovarian aging matters?

Ovary is the first organ to undergo functional decline

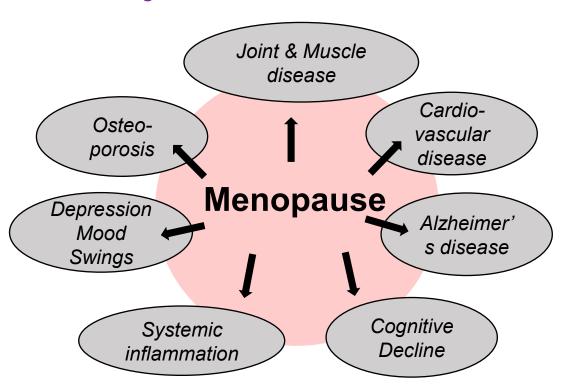
Fertility by age

Fertility

Menopause

20-24 25-29 30-34 35-38 40-44 45-48 years

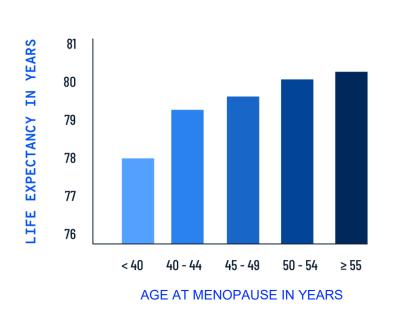
Menopause marks a systemic shift, increasing risks for various chronic conditions



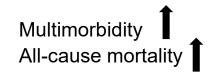
Menopause, the endpoint of ovarian aging, marks the onset of accelerated biological aging and chronic disease risk

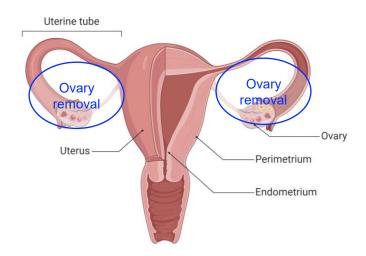
Ovarian function influences systemic aging in women

Women with later age of menopause live longer

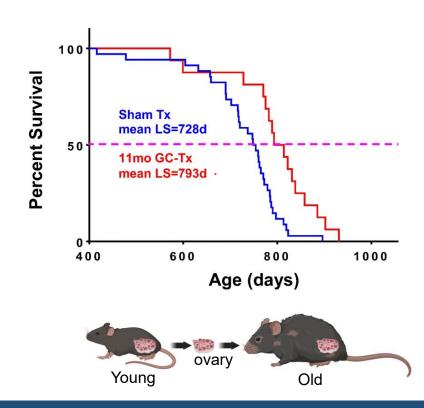


Negative impacts of oophorectomy in women





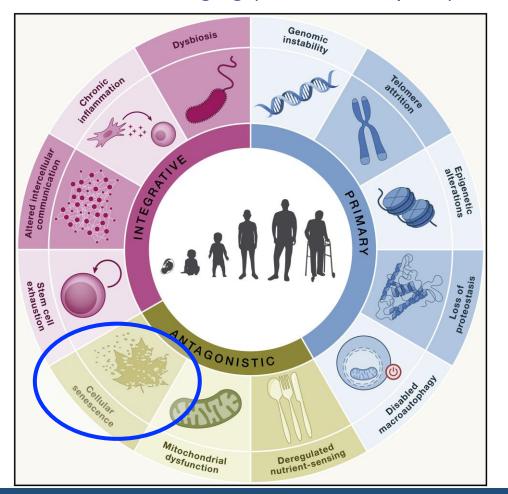
Transplanting young ovaries extends lifespan in aged mice

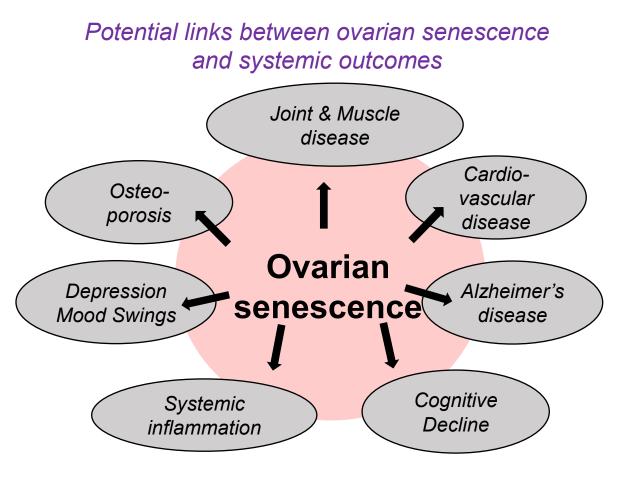




Cellular senescence: One of the biological processes behind ovarian aging

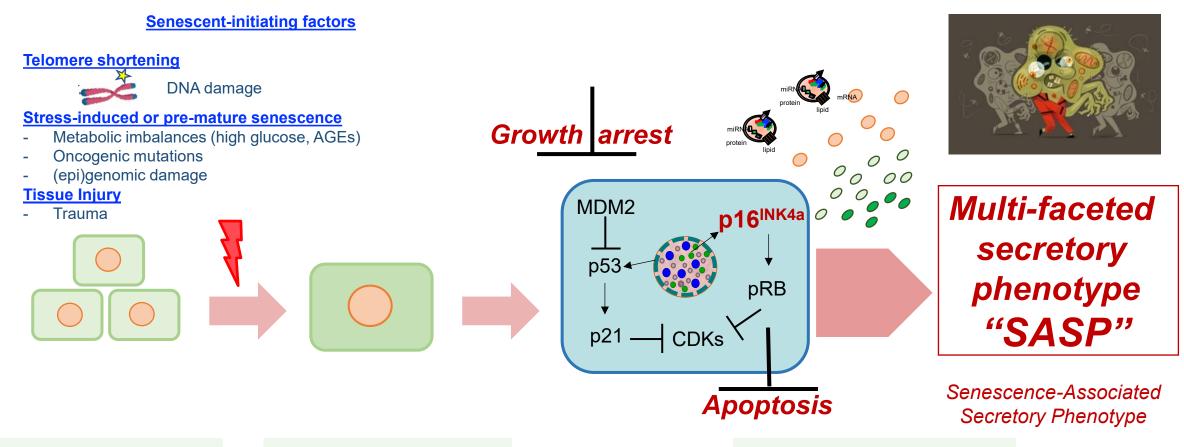
13 hallmarks of aging (+ECM discruption)





What is cellular senescence?

Aging doesn't just happen at the tissue or organ level. It starts with changes inside individual cells

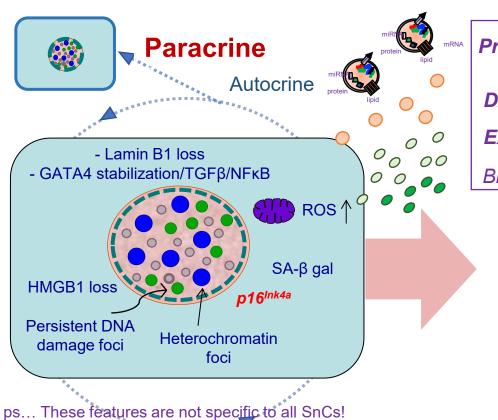


Proliferating cells (non-senescent cells, non-SnC)

Initiation of senescence

Stabilization of senescence (senescent cells (SnC))

How senescent cells drive aging and diseases?



Pro-inflammatory (cytokines, chemokines, growth factors, proteases)

DAMP (HMGB1 loss/secretion, cf-mt-DNA)

Extracellular Vesicles (MVs & exosomes)

Bioactive lipids, miRNAs, microsomes etc

Inflammation + (Inflamm-aging)

Perturbed signaling

Destroy tissues (ECM disruption)

Disrupt normal cell functions

Prevent stem cell functions

Our question:
Ovarian to
Systemic aging
in women

Diseased tissues

Accumulation of SnC

Not all SnCs express these markers

Spread of SnC and "sterile" Inflammation

When and where do senescent cells occur?

With increasing age

Human, non-human primates, rodents, etc





At sites of age-related pathology

- Neurodegenerative diseases (Senescent neurons & glial cells)
- Eye degenerative diseases (Senescent RPE, lens, retinal cells)
- Pulmonary fibrosis (Senescent pulmonary cells)
- Cardiac fibrosis (Senescent cardiomyocytes)
- Cancer metastasis and recurrence (Senescent cancer cells)
- Degenerative disc diseases (Senescent NP/AF cells)
- Liver fibrosis (Senescent liver cells)
- Kidney fibrosis (Senescent kidney cells)
- Ovarian diseases/Infertility (Senescent granulosa cells)
- Diabetes (Senescent β-cells)
- Osteoporosis (Senescent osteoblasts and osteocytes)
- Osteoarthritis (Senescent chondrocytes)
- Sarcopenia/frailty (Senescent satellite cells and fibro-adipogenic progenitors)

Embryogenesis, timely parturition, wound healing placental development during pregnancy

NOT ALL BAD!

Senotherapies: from joints to ovaries

- Senolytics = a new class of drugs that 'selectively' kill senescent cells
- Senomorphics = drugs that 'selectively' suppress certain SASP modules





Osteoarthritis

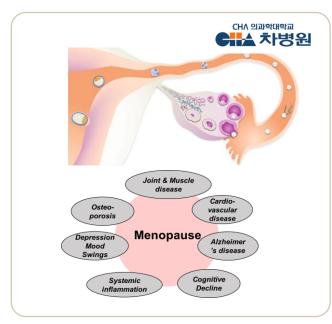
Clinical trial phase 2b (NCT0412994);
Patents 10,130,628; 9,993,472; 9,855,266; 9,849,128

Jeon et al., Nat Med, 2017; Jeon et al., JCI, 2018; Jeon et al., JCI insight, 2019; Faust et al., JCI; Gil et al., Aging, 2022; Chin et al., Aging cell, 2023



Muscle injury / Sarcopenia

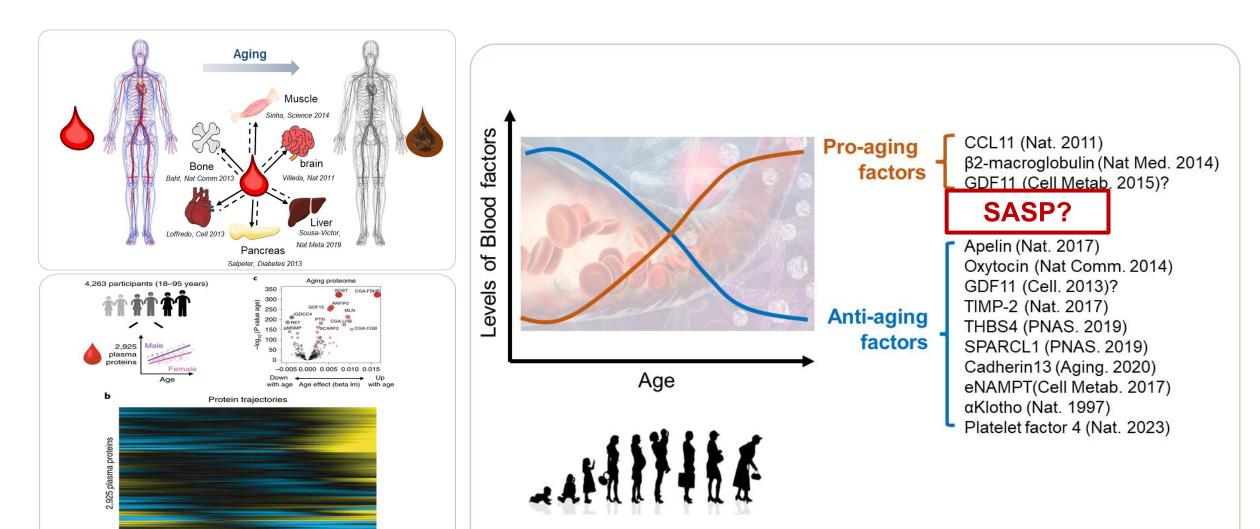
Jeon et al., Nat Met, 2022; Kim and Gil et al., Journal of Cachexia, Sarcopenia and Muscle, 2025



Ovarian Aging

Shin et al., Rejuvenative Research, 2022; Shin et al., Menopause, 2023; Gwak et al., Envir Res, 2024

Blood-borne factors reflect biological aging



Age (years)

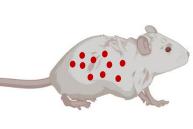
zscore

Systemic circulation of SASPs may link aging and rejuvenation

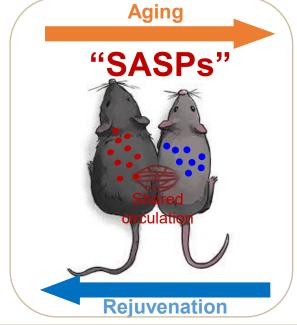
"Heterochronic Parabiosis"

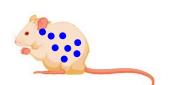
Pro-aging factors

- Scare in young animals
- Increase with age
- Reduce tissue homeostasis in young animals



18-mon-old mouse (≈ 65-yr-old human)





3-mon-old mouse (≈ 20-yr-old human)

Muscle (Conboy, Nat 2005; Sinha, Science 2014)

Brain (Villeda, Nat 2011, 2014; Yousef 2019; Kim, IJMS 2020),

Liver (Conboy, Nat 2005; Sousa-Victor, Nat Meta 2019)

Pancreas (Salpeter, Diabetes 2013)

Heart (Loffredo. Cell 2013)

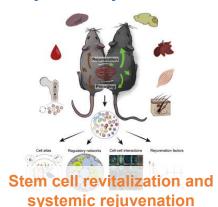
Bone (Baht, Nat Comm 2013; Yi, Swiss Med Wkly

2018; Li Metabolites 2020)

Anti-aging factors

- Abundant in young animals
- Decline with age
- Rejuvenate tissues (heart, brain, muscle, liver, bone, pancreas, kidney, liver, aorta, intervertebral disc) when supplied to old animal

Stem cell revitalization and systemic rejuvenation







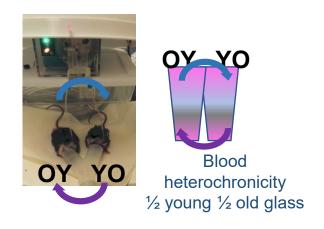
Adult stem cell rejuvenation

factors

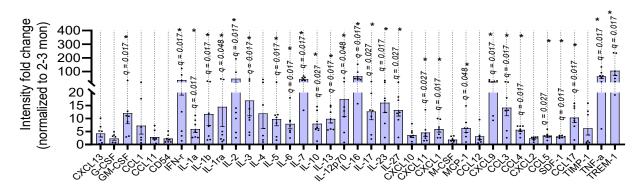
Ma et al., Cell Stem Cell, 2022

Do circulating SASPs spread aging?

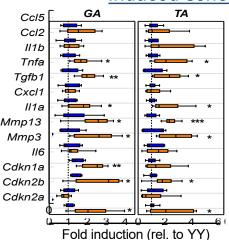
Heterochronic apheresis

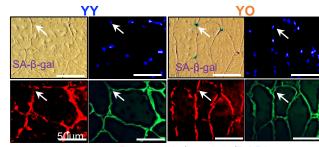


Circulating SASP factors in the old blood



Induced senescence in young skeletal muscle

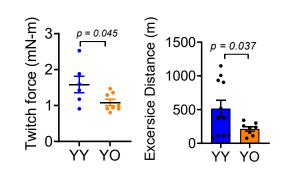


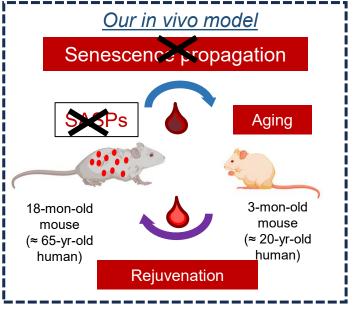


Dystrophin/Laminin/DAPI

* SA-β-gal+ satellite cells that were located between the laminin+ basal lamina and dystrophin-outlined subsarcolemma

Functional tests

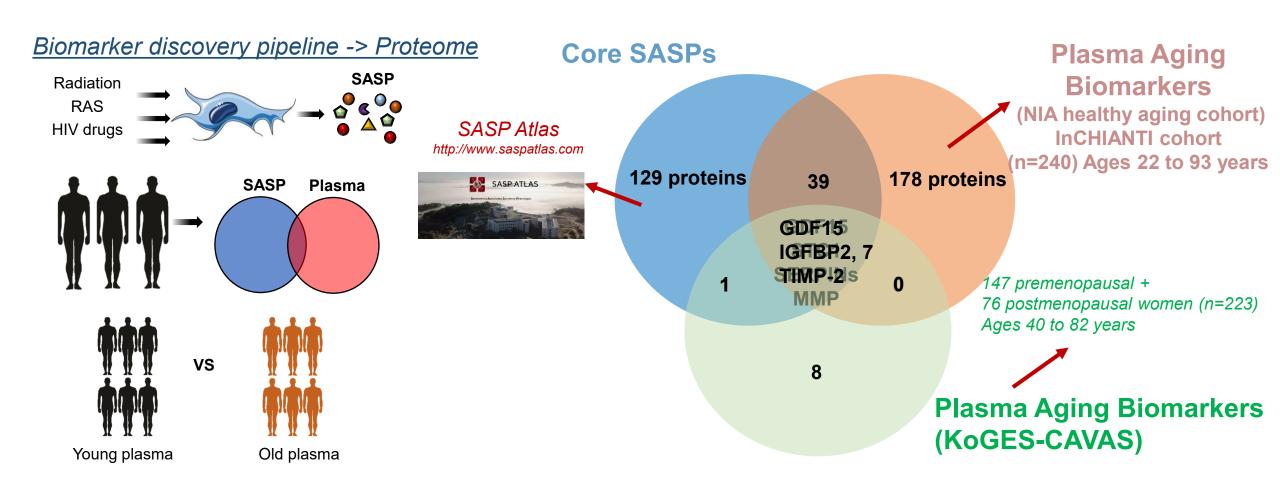




Jeon et al., Nature Metabolism, 2022

Core SASPs overlap with plasma aging markers in human cohorts

SASPs are in aged human blood



Do circulating SASPs reflect systemic signals of ovarian aging, particularly in postmenopausal women?

SASP and ovarian aging in a prospective population-based cohort study

Study participants

- 147 pre- $(46.4 \pm 3.9 \text{ y})$ and 76 $(67.0 \pm 6.9 \text{ y})$ post-menopausal women in the Korean Genome and Epidemiology Study Cardiovascular Disease Association Study (KoGES-CAVAS), a population-based prospective cohort study conducted by Korea NIH

Measured markers

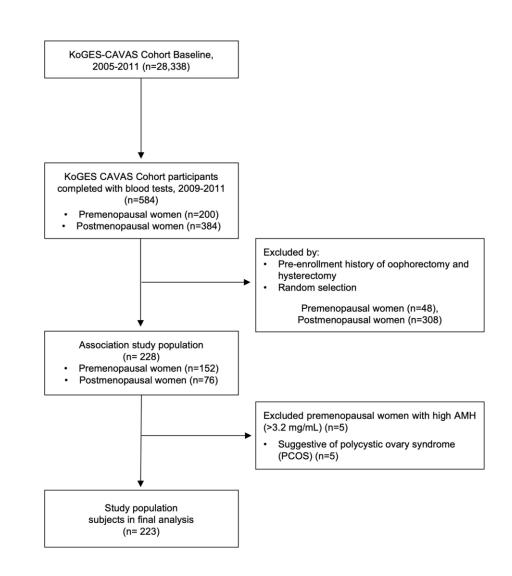
- 32 SASP proteins overlapped with aging-related markers and AMH in human plasma

Outcomes

- Chronological age
- **Biological age:** menopausal state (menopausal age and years since menopause) and AMH level (mean 0.5 ± 0.7ng/ml)

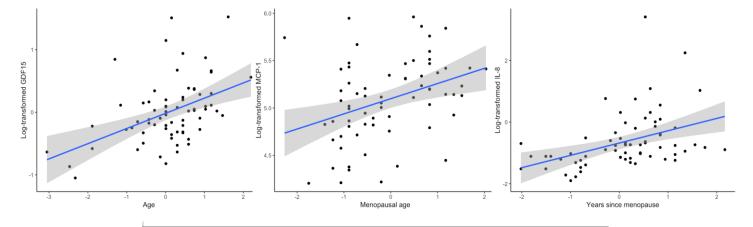
Statistical methods

- Multivariate analysis adjusted for age and BMI



Plasma SASPs reflects menopausal and chronological ages

- Investigated circulating 32 SASP proteins in 76 post-menopausal women (avg 67 ± 6.6 y; range 46–82 y) from KoGES-CAVAS study
- Identified GDF-15, IGFBP-2, and TNFα as correlates of chronological age
- IL-8 and MCP-1 were associated with menopausal age and years since menopause
- A combination of 13 SASPs proteins are selected to be associated with chronological, menopausal age and years since menopause using multiproteins model



Variables	Chronological age				
	Adjusted for menopausal age	Adjusted for years since menopause	Menopausal age	Years since menopause	
GDF15	0.400	0.141	-	-	
IFN-γ		-	-0.019	-	
IGFBP-2	0.213	0.053	_	-	
IGFBP-7	0.284	-	_	-	
IL-1β	-	-0.250	-0.218	0.155	
IL-15	-	-	-0.025	0.004	
IL-17A	0.153	0.040	_	-	
IL-8	0.020	-	-0.132	0.090	
MCP-1	-	0.225	0.495	-0.336	
TIMP-2	-	-0.120	-0.349	0.169	
TNF-α	0.113	-	-0.120	0.046	
VEGF-A	-	-0.021	-0.045	0.023	
IP-10	0.006	0.104	0.048	-	

SASPs can act as systemic indicators of ovarian aging, reflecting senescence burden that evolves with ovarian aging

Association of SASP proteins with menopausal status

SASP	Coefficient (SE)	<i>p</i> -value	Adjusted <i>p</i> -value ^a
Eotaxin	0.457 (0.126)	<0.001	0.001
GDF15	-0.643 (0.266)	0.017	0.030
IFN-γ	0.544 (0.165)	0.001	0.003
IGFBP-2	6.838 (0.181)	<0.001	<0.001
IGFBP-4	7.038 (0.145)	<0.001	<0.001
IGFBP-5	7.057 (0.238)	<0.001	<0.001
IGFBP-7	6.940 (0.068)	<0.001	<0.001
IL-1α	-1.047 (0.392)	0.009	0.017
IL-2	-0.414(0.145)	0.004	0.009
IL-6	-0.757 (0.282)	0.008	0.016
IL-7	-0.567 (0.139)	<0.001	<0.001
IL-8	-0.597 (0.219)	0.007	0.015
IL-10	-0.354 (0.154)	0.022	0.036
MCP-1	0.821 (0.153)	<0.001	<0.001
MIP1α	-0.628 (0.208)	0.003	0.008
TIMP-1	0.378 (0.105)	<0.001	<0.001
TIMP-2	0.249 (0.078)	0.001	0.003

^{*} Pearson correlation coefficient (SE) was calculated by linear association using generalized linear regression model. Adjusted p-value for the false discovery rate derived with the Benjamini-Hochberg method.

^{**} Blue boxes indicate SASPs that are positively associated with chronological age

Association of SASP proteins with AMH level

SASP	Coefficient (SE)	<i>p</i> -value	Adjusted <i>p-</i> value ^a
IL-6	0.186 (0.082)	0.026	0.560
IFN-y	0.075 (0.036)	0.040	0.560
VEGF-A	0.112 (0.077)	0.147	0.714
IGFBP-7	-0.015 (0.011)	0.176	0.714
GDF15	-0.093 (0.075)	0.212	0.714
TIMP-2	0.112(0.077)	0.255	0.714

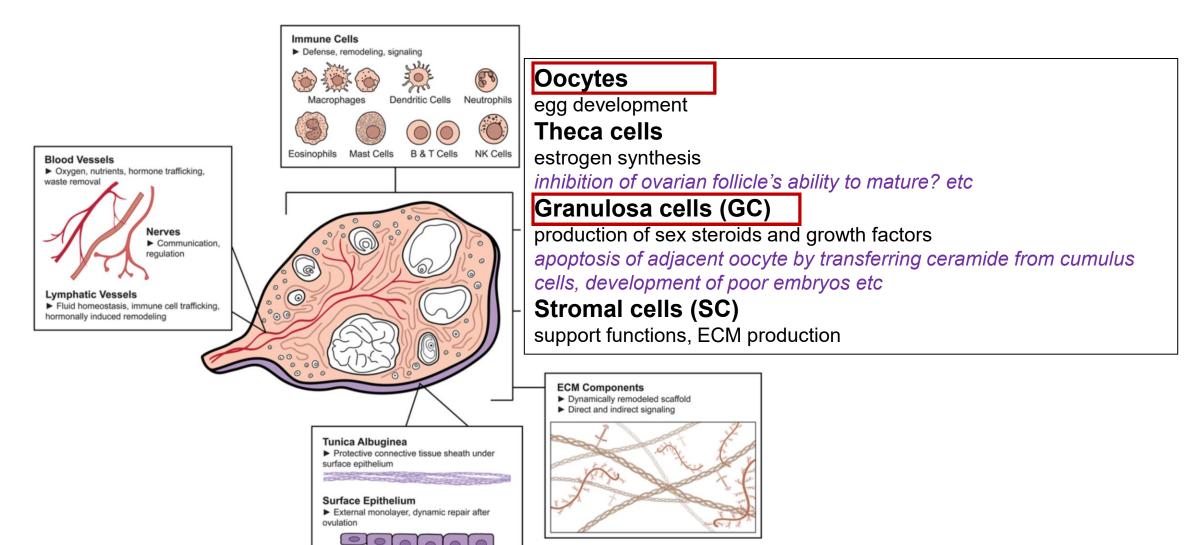
^{*}Pearson correlation coefficient values (standard error) were calculated by linear association using generalized linear regression (GLM) model. Adjusted p-value for the false discovery rate derived with the Benjamini–Hochberg method.

- No significant associations between SASP levels and AMH in premenopausal middleaged/older women
- Subgroup analysis (<45 y) showed lack of association: lack of younger participants?

Plasma SASPs reflect menopausal status but may not be reliable biomarkers of ovarian reserve in midlife women

^{**} Blue boxes indicate SASPs that are positively associated with chronological age

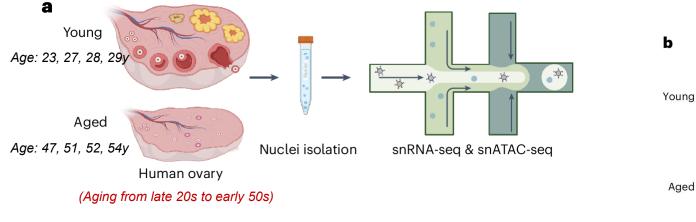
Complex biology of human ovarian aging and senescence

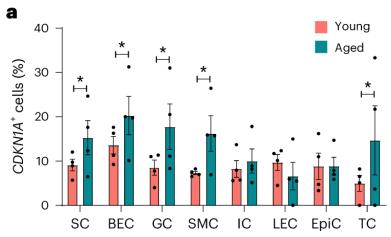


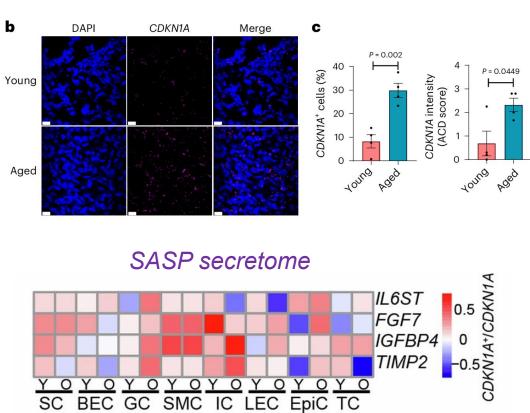
Ovary is composed of various cell types, each potentially contributing differently to cellular senescence and aging

Which cells become senescence in human aged ovary?

Increased proportion of senescent cells (p21+ cells) in the aged ovary



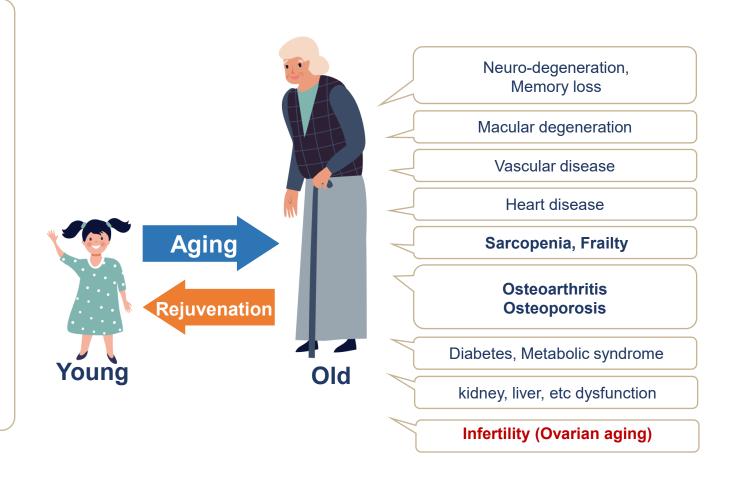




Aging can be modified by anti-aging interventions

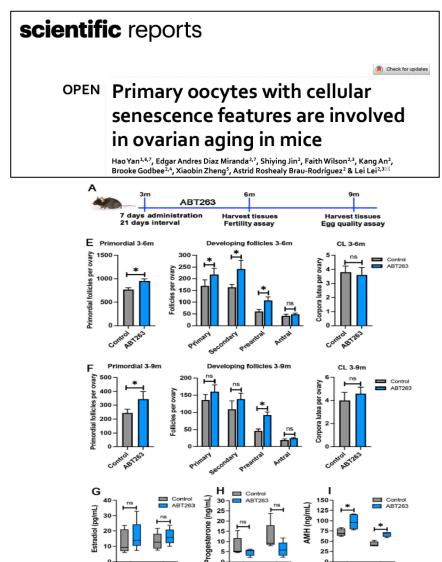
Rejuvenation —not just slowing aging but actually reversing it — is feasible at least in mice!

- Epigenetic reprogramming of aged cells
- Metabolic interventions (dietary restriction, exercise, etc.)
- Elimination of harmful senescent cells
 : Senotherapeutics (senolytics & senomorphics)
- Utilization of factors within bloodstreamYoung blood plasma treatment

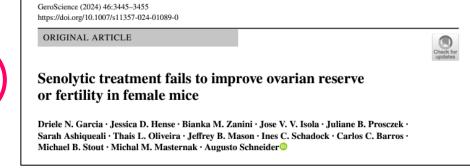


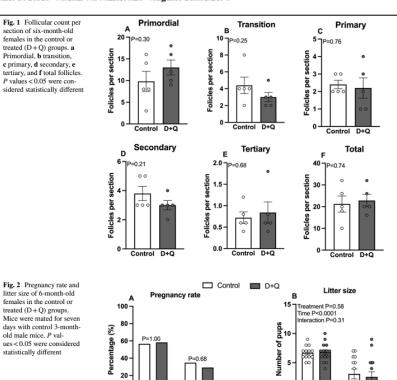
Senotherapy shows variable efficacy in delaying ovarian aging in mice











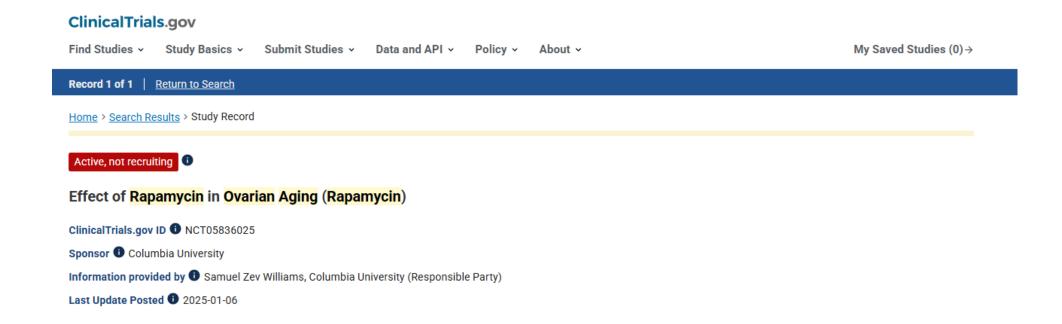
Better understanding of ovarian senescence is needed for clinical translation

Clinical trial of Rapamycin for delaying ovarian aging in human

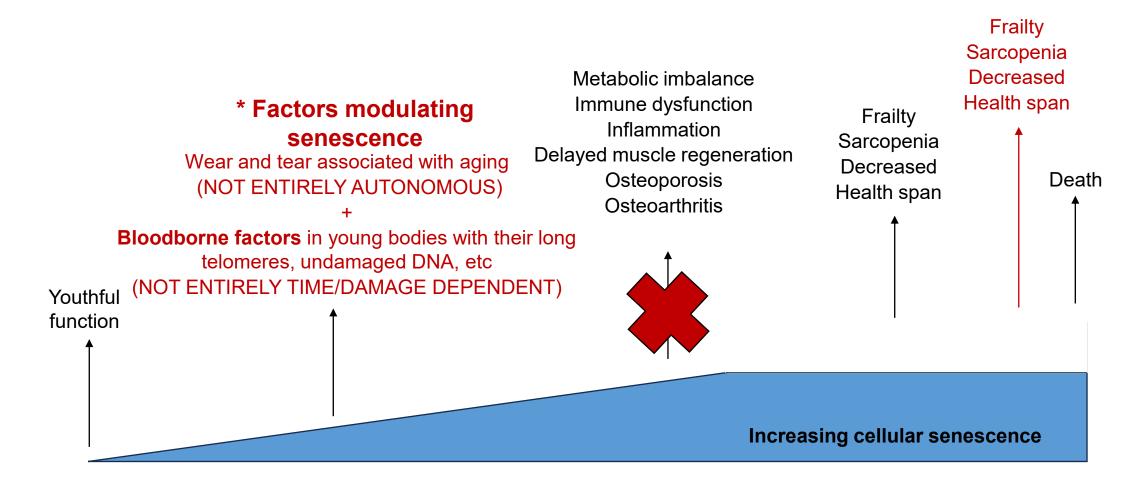
A randomized, double-blind, placebo-controlled pilot study

Population: Premenopausal women (age 35-45y)

Primary Endpoint: Ovarian function and aging biomarkers (e.g., AMH levels, follicle count)



Key Takeaway: Targeting senescence to treat and prevent agerelated diseases



Our lab focus: developing senotherapies for age-related disease

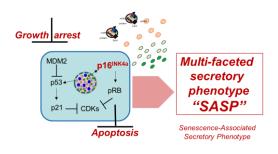
Cellular senescence are key factors in impairing regeneration in response to injury and aging

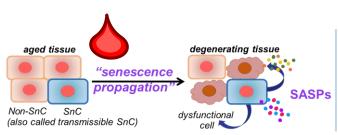


Senescent cells trigger secondary aging in neighboring cells, drive systemic aging, and alter tissue microenvironment



Develop senotherapies to create a pro-regenerative environment and treat age-related diseases







Map senescence in musculoskeletal diseases and others & Discover associated biomarkers (scRNA-seq, spatial transcriptomics, proteomics)

Define senescence's impact on young cells and tissue function (2D, 3D bioprinted and microfluidics based aged tissue model, in vivo, patients)

Develop rejuvenation intervention to target senescent cells (e.g., senolytics)

Senescence is a treatable hallmark, and our lab develops targeted therapies to reverse aging-related degeneration

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Geun Hyung Kim (SKKU)









Thank you for your attention!

Baseline characteristics of participants

	Premenopausa	al women (n=147)	Postmenopaus	sal women (n=76)	
Variables	Mean ± SD ^a	Median (IQR) ^b	$Mean \pm SD$	Median (IQR)	Р
Age (years)	46.4 ± 3.9	46 (43–50)	66.9 ± 6.9	68 (63.5–71)	<0.001
BMI (kg/m²)	23.9 ± 3.2	23.7 (21.7–25.7)	23.5 ± 3.0	23.3 (21.1–25.3)	0.536
AMH (ng/mL)	0.5 ± 0.7	0.2 (0.1–0.7)	_	<u>-</u>	_

All of ± data indicates mean ± standard deviation of the mean. Median (interquartile range) of the level of SASP in study population. P-values were analyzed by a two-tailed unpaired t-test or the Mann–Whitney U test was used to compare means or medians, respectively, of the SASP according to menopausal status

^{*} Year since menopause: This variable is often used as an indicator of cumulative biological aging or hormonal deprivation over time